THE WELFARE OF BROILERS IN THE EU



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INTRODUCTION

The EU is a major producer of chickens reared for meat (known by the industry as broilers), being responsible for 8.6% of total world production (10.6 million tonnes)¹. Broiler production in the EU has increased by around 20% from 2012 to 2022, now representing about 6.1 billion birds every year¹. Production and consumption have been increasing steadily and with an average consumption of 23.4 kg per capita per year in 2022, chicken comes second after pig meat as the highest consumed meat in the EU². Poultry meat production in the EU is expected to increase by 0.2% and consumption by 3% in the period 2022-2032, whereas pig meat and beef are expected to keep declining². The number of broilers slaughtered in the EU is around 25 times higher than pigs³, the second most slaughtered land animal in the EU.

Intensive broiler systems account for the vast majority (around 90%) of broiler production in the EU⁴, while alternative systems (alternative indoors, free-range, and organic) represent a small proportion of total production (less than 10%). The broiler industry is extremely concentrated; while there are over 20,000 professional broiler producers, the top 10 alone account for over 40% of broiler production⁵. Three-quarters of the EU production is concentrated in seven Member States: Poland, Spain, France, the Netherlands, Germany, Italy, and The Netherlands, which are also home to the largest producers (figure 1).

FIGURE 1: CHICKEN MEAT PRODUCTION BY EACH EU 27 MEMBER STATE ¹



Share of EU 27 chicken meat production, 2022



GENETIC SELECTION

The number of broilers slaughtered in the EU is higher than ever, and the broilers themselves grow faster every year. The chickens produced today are genetically selected to grow as big as possible, as quickly as possible, using as little feed as possible. While there may be economic advantages to this, it comes at a significant cost to bird health and welfare. The genetic selection of broilers has led to a 400% increase in broiler growth rate, achieving market weight in 60% less time than broilers 50 years ago. The amount of breast meat on an individual bird increased by two-thirds⁶. These broilers achieve the target live weight of 2-2.5 kg in around 34-39 days, in comparison to 16 weeks back in the 1950s^{7,8}. The vast majority of the EU chicken meat sector uses these fast-growing strains. The most widely used breed in all Member States is the Ross 308, which accounts for at least 70% of all broilers slaughtered in the EU, while Hubbard and Cobb breeds are less widely used⁹. Achieving the best food conversion rate with the premium cut of breast meat is the economic driver for the broiler industry we have today. The sentience of the animal is not part of the economic program. However, as a recent paper shows, chickens are cognitively intelligent, they can demonstrate self-control, they communicate with one another in a complex way, and have the capacity for reason and logic¹⁰.

POOR LEG HEALTH

One of the most serious welfare problems in broiler production is the high incidence of skeletal disorders, particularly those that lead to impaired mobility or lameness¹¹. The development of many of these conditions is related to selection and management for rapid growth, since they are rarely seen in slower growing strains and laying strains of poultry but are very common in commercial fast-growing chickens such as the Ross 500. Skeletal problems are not just a welfare issue; they are also costly to the industry. They are by far the most costly diseases for poultry producers in terms of output loss, resource wastage, and treatment and prevention costs¹².

The development of large muscle mass on an immature skeleton leads to locomotor problems in fast-growing chickens¹³, with the highest levels of lameness seen in the fastest growing birds¹⁴. Even broilers with moderate lameness (\leq gait score 3 on a score from 0 - 5) have been shown to suffer pain from their impaired walking ability¹⁵. Chickens given the ability to self-medicate with feed containing painkillers consumed a significantly higher proportion of this feed as the severity of lameness increased¹⁶. Around 27.6% of commercial intensively-reared broilers presented leg abnormalities and 3.3% were almost unable to walk, according to a study from the UK¹⁷.



Poor leg health can be caused by a number of different factors, including bacterial chondronecrosis and osteomyelitis (BCO), sometimes referred to as femoral head necrosis^{14,18,19}, tibial dyschondroplasia^{20,21}, and viral arthritis^{22,23}. In severe cases, birds lose the ability to walk so they can't even access the basic resources of food and water, which can lead to starvation and a slow and painful death if they are not culled. Broilers are typically reared in barns with a stocking density of around 39 - 42 kg/m² (19.5 - 21 birds/m² when slaughtered at 2 kg). Such high stocking densities negatively affect their walking ability¹⁷. Fast-growing broilers with poor leg health spend an increased amount of time sitting, and long periods of time spent on poor litter which can lead to the painful condition footpad dermatitis (inflammation and necrotic lesions on the plantar foot), as well as hock burns and breast blisters^{24,25}. The ability to walk is absolutely necessary to avoid pecking from other aggressive birds in the flock²⁶.

ASCITES

Ascites, commonly known as 'water belly', is an accumulation of fluid in the abdominal cavity²⁷. Since ascites develops gradually, the birds suffer for an extended period before they die²⁸. It is a multifactorial disorder. However, the main contributor of the condition is believed to be an increased oxygen demand by the fast-growing muscle. The increase in blood pressure required to push the blood through the blood capillaries in the lung and the increase in workload for the right side of the heart results in pulmonary hypertension and ventricular hypertrophy. The increased blood pressure in the veins, liver, and abdominal vessels forces plasma fluid out of the vessels, particularly the ones of the liver, into the abdominal space^{26,29,30}. Thus, the increase in metabolic demand, coupled with exposure to environmental conditions such as temperature, lighting and ventilation, and nutritional factors such as feed form or content, all seem to promote the development of ascites^{31,32}. It is most commonly seen in male chickens and ascites mortality can range from 0 to 30% in broiler flocks³³. It is the main cause of carcass condemnations in UK slaughterhouses since 2003 (reaching an annual high of 2.7 million birds in 2013)³⁴. In a study conducted in Norway, ascites was found to be the most common condemnation cause of fast-growing broilers sent to slaughter. The frequency of ascites in fast growing broilers was seven times higher than in slow-growing broilers (0.661% and 0.093% respectively)³⁵.



SUDDEN DEATH SYNDROME

Sudden death syndrome (SDS), or 'flip-over', is a condition in which apparently healthy fast-growing broilers die unexpectedly from no apparent causes³⁶. Birds will suddenly start violently flapping their wings, extend their neck, squawk and die within minutes³⁷. The condition is estimated to cause mortality of 0.8 to 4% in broiler flocks, with males predominantly affected³⁸⁻⁴⁰.

The cause of flip-over in broilers is still unknown. It is often associated with nutrition (high density diets)⁴¹ and environmental factors (noise, lighting)³⁶, but a growing body of evidence suggests that it may relate to broilers' high predisposition to cardiac arrhythmia⁴². Ventricular fibrillation appears to be the immediate cause of death⁴³. Sudden death syndrome can occur as early as 2 days of age and continues until birds reach market weight. Peak mortality usually occurs between days 21 and 27⁴⁴.

BREAST MUSCLE MYOPATHIES

Muscle myopathies, or diseases of the muscle tissue, impact both bird welfare and meat quality. They are more commonly seen today than in previous years and are all associated with the intensive genetic selection of broilers to improve their growth rate, body weight, and breast yield⁴⁵⁻⁴⁹. Deep pectoral muscle myopathy has been studied for some time, while other abnormalities such as white striping, wooden breast and spaghetti meat have not been reported until recently^{50,51}.

Deep pectoral myopathy (DPM) is caused by a lack of blood supply to the pectoralis minor, known as the mini filet to consumers, leading to necrosis of the tissue. This is caused by the overgrown pectoralis major actually compressing the pectoralis minor. This means that when the bird flaps its wings the pectoralis minor is unable to receive the oxygen and nutrients that it needs⁵². It is known as green muscle disease due to the appearance of green flesh within the muscle tissue⁵³. The occurrence of DPM in broilers is estimated to vary between 0.02% and 1.9%^{54,55} with more cases reported in faster growing strains and in males⁵⁶.

Wooden breast (WB) is characterised by necrotic muscle fibres and the replacement of muscle with connective tissue, water, and fat, causing a palpably firm consistency of the breast muscles⁵⁷. WB often occurs with the condition known as 'white striping' (WS)⁵⁸. WS is found on the outside of the pectoral major muscle. It is visible as white striations running parallel to the muscle fibres⁵⁹. These striations are found to be



adipose (fat) tissue⁶⁰. Even though the cause of both conditions is unclear, several studies have suggested that their pathogenesis is associated with several biological processes, such as localised hypoxia within the muscle, oxidative stress, increased intracellular calcium build-up, and repair of cellular damages^{50,61–63}.

Although the incidence rate of WB in commercial chickens is not well known, it is becoming increasingly common⁶² and flocks that are affected have up to 50% of birds with the disease^{58,64}. Affected chickens are most likely to be those that grow faster, have greatest feed efficiency, heaviest body weight and higher breast muscle yield^{47,49}. A recent study found that the incidence of WB in slower growing birds was less than 1%, compared to 3.7 - 23.4% in the three faster growing breeds tested⁶⁵. Another study found that WB is associated with an impairment of gait scores, and may thus be partly linked to the common walking abnormalities in broilers⁴⁹.

While WB is apparently asymptomatic, clinical signs such as outbulging of the lateral forebreast and decreased wing movement have been noted in severely affected birds. Also, the degenerative process leading to WB is similar to Duchenne muscular dystrophy in humans⁵⁷, a painful and debilitating condition^{66,67}. Thus, it may be possible that broilers affected with WB also experience similar pain and discomfort.

Meat that comes from birds suffering from WB or from those with both conditions are found to have a harder texture, impaired ability to hold water, and poorer nutritional value⁶⁸. WS has been found to increase the fat content and decrease the protein content of affected fillets⁶⁹ and also impacts the general appearance of the breast meat⁵⁹. These conditions are forcing the downgrading of meat due to the lack of aesthetic appeal⁴⁷ and it is estimated that the incidence of these conditions can result in an excess of \$200 million (€176 million) per year lost in the US⁵¹. It's estimated that the disposal of breast muscles by poultry processing plants in Poland due to DPM causes annual losses of €2 million⁵⁴.

More recently, a new muscular abnormality termed as 'spaghetti meat' (SM) has emerged⁷⁰. SM, often associated with WS, is characterised by poor muscle cohesiveness due to the immature intramuscular connective tissues. The affected muscle is so loose in structure that the muscle fibre bundles can be pulled away easily with the fingers, like spaghetti^{48,50}. Broilers displaying higher breast size seem to be more prone to be affected by SM⁷⁰.



STOCKING DENSITY

Stocking density is considered one of the most important factors for the welfare of broilers. Although there is no doubt that keeping broilers at high stocking densities compromises health and welfare, birds continue to be given very little space to move around with varying degrees of density allowed by EU law and in specific countries.

The EU Broiler Council Directive 2007/43/EC outlines the minimum required environmental conditions for maximum stocking densities. According to the Directive 2007/43/EC, the maximum stocking density in a holding or a poultry house on a holding should not at any time exceed 33 kg/m². A higher stocking density of a maximum of 39 kg/m² is permitted with a set of environmental requirements. The stocking density may rise to a maximum of 42 kg/m² if cumulative daily mortality rate is low. Environmental conditions include ventilation, heating and cooling systems to maintain the appropriate temperature, humidity and CO₂ and NH₃ concentrations⁷¹.

However, some Member States have chosen to go beyond these requirements by implementing more stringent legislation or standards. Maximum stocking densities have been set in Austria (30 kg/m^2), Denmark (40 kg/m^2), Sweden (36 kg/m^2), Germany (39 kg/m^2), and the UK (39 kg/m^2)⁹.

High stocking density in broiler sheds restricts the chickens' behaviour and causes health problems. Studies have shown that higher stocking densities decrease locomotor activity, stretching behaviour, walking, eating, preening, and shaking^{72,73}. Jostling of other birds, disturbance of resting birds, birds climbing on top of one another^{74,75}, and fights⁷⁶ are also observed at higher stocking densities. The restriction of space and locomotor activity in crowded sheds can reduce the consumption of feed, which is followed by a decrease in final body weight^{77–79}. Carcass quality may also be compromised due to scratches, bruising, poorer feathering, and condemnations ^{80,81}.

High stocking density leads to greater litter moisture, increased microbial activity, and increased temperature and ammonia concentration which can give rise to hock burn, foot-pad dermatitis, breast blisters ^{24,28,79,81–84} and respiratory diseases⁸⁵. High levels of ammonia can compromise their immune system, increasing the birds' vulnerability to infections⁸⁶.



LIGHT

The majority of broilers produced in the EU are reared in environmentally controlled buildings without windows, where artificial light is provided. Light is an important factor as it allows the bird to establish rhythmicity and synchronise many essential functions like feeding and digestion, body temperature and reproduction⁸⁷. Under natural conditions, birds are active during the daytime light period and rest and sleep at night when it is dark. There is an increase in activity around dawn and dusk, as the birds forage for food. Long day lengths can cause sleep deprivation, which can negatively impact broiler welfare⁸⁸. Despite this knowledge, some global producers routinely utilise almost continuous light regimes, at low light intensities, with the notion that these lighting schedules decrease activity and increase feed intake, consequently maximising growth rate and production.

According to the Council Directive 2007/43/EC, all poultry buildings should have lighting with an intensity of at least 20 lux during the lighting periods, measured at bird eye level and illuminating at least 80% of the usable area. Lighting must follow a 24-hour rhythm and include periods of darkness lasting at least six hours in total, with at least one uninterrupted period of darkness of at least four hours from 7 days of age onwards until three days before slaughter⁸⁹.

Low light intensity has a negative effect on broilers' welfare as it impacts activity level, behaviour patterns, performance of comfort behaviours, foot and eye health^{90–97}. Broilers perform more active behaviour, such as foraging, under the brightest light available (200 lux)⁹⁸. It has been shown that lighting programs with a minimum intensity of 50 lux stimulate higher diurnal activity levels in broilers without negatively affecting weight gain⁹⁹.

Continuous lighting is detrimental to broilers as it has been shown to decrease activity and comfort behaviours such as preening and wing-shaking^{100,101}. Fearfulness (shown by a behavioural measure) is greater in broilers reared under continuous light compared to those reared with a proper dark period^{100,102-104}. Normal ocular development in the chick requires a minimum of four hours of darkness per day, provided at the same time of the day without interruption¹⁰⁵. Constant lighting results in the disruption of behavioural rhythms for broiler flocks, and four hours of darkness may not be enough to ensure full rhythm development⁸⁸.



Research has shown that giving broilers an uninterrupted dark period resulted in decreased mortality¹⁰⁶, leg¹⁰² and foot problems¹⁰¹, rates of sudden death syndrome^{36,107} and ascites¹⁰⁸, and improved broilers' antioxidant status and nonspecific immunity¹⁰⁹. Given these clear impacts on health and welfare, current dark periods should be increased from the standard four hours to a minimum of six hours and must be continuous.

Providing natural light is extremely important as it reduces the percentage of time that broilers spend lying and improves leg health¹¹⁰. Chickens spend more time drinking, exploring, moving, and foraging in natural light compared to artificial lighting¹¹¹. The minimum requirement of 50 lux ensures that sufficient windows are installed to provide this. The table below shows common indoor and outdoor light levels, demonstrating the ease at which 50 lux should be able to be met within broiler sheds. In the situation where 50 lux cannot be met through natural light alone, artificial lighting can be used to compensate.

TABLE 1 - COMMON OUTDOOR AND INDOOR LIGHT LEVELS¹¹²

Condition	Illumination (lux)	
Sunlight	100,000+	
Full Daylight	10,000+	
Overcast Day	1000+	
Very dark Day	1000+	
Classrooms	300	
Libraries	300	
Entrance Hall	200	
Stairs	150	



ENVIRONMENTAL ENRICHMENT

Environmental enrichment is defined as "an improvement of the environment of captive animals which increases the behavioural opportunities of the animal and leads to improvements in biological function"¹¹³. Broiler houses are usually barren environments, which contribute to the low behavioural activities of broilers. The addition of enrichment items has been shown to increase activity levels¹¹⁴ and the expression of comfort behaviours¹¹⁵. Interestingly, birds in enriched houses show higher levels of several activities even in areas where no enrichments are present¹¹⁶.

For slow-growing breeds with outdoor access, providing access to perches inside the house increased the percentage of time the birds spent standing¹¹⁷. Provision of perches has been associated with the reduction of disturbances (pushing and trampling)¹¹⁸ and in the number of hock burns and foot pad dermatitis^{119,120}. Broilers have been observed to use perches from as early as six days of age, and on average from nine days of age¹²¹. Platforms have been found to positively affect leg health, as birds with access to platforms have improved gait scores and lower prevalence and severity of tibial dyschondroplasia¹²¹. Provision of panels has been observed to reduce disturbances during rest¹²² and also serve as shelter areas¹²³.

Provision of straw bales and pecking objects, such as bundles of string, in environments with natural light affect walking ability and decrease time spent lying down^{110,124}. Provision of multiple enrichments results in higher overall activity levels and a higher likelihood of birds engaging with the enrichment items¹²⁴.

SLAUGHTER

In Europe, the welfare of animals—including poultry—at the time of killing is protected under Regulation (EC) 1099/2009. This regulation requires the use of approved stunning methods for poultry in the EU and extends to slaughterhouses in third countries that export meat to the EU. Today 53% of broilers in the EU are slaughtered in an electrical water-bath system¹²⁵. With this method of slaughter, conscious birds are hung by the legs upside-down on a moving metal shackle line and their heads pass through an electrified water-bath before having their throats cut¹²⁶.

Water-bath stunning was created to allow fast processing of birds, however there are many welfare problems associated with this stunning system. The birds' legs are compressed during shackling, causing pain¹²⁷, especially in birds with thicker legs or



suffering from painful lameness due to leg diseases, bone dislocations or fractures. Bird inversion increases the levels of stress that poultry are subjected to during the shackling process¹²⁸. Rough shackling can significantly contribute to wing flapping leading to dislocations and bone breakages. Pre-stun electric shocks can occur if the birds' wings make contact with the water-bath before their heads do^{129–131}. It has been shown that electrical stunning is not completely effective. Occasionally some birds are not properly stunned because they miss the stunner by raising their heads and missing the water¹³², or when their heads do enter the water-bath but the currents are too low to induce unconsciousness^{126,133}.

Controlled atmosphere stunning (CAS), or gas stunning, has become increasingly common during the last 20 years in Northern Europe, mainly as a result of the animal welfare and product quality advantages in comparison with water-bath stunning¹³¹. The percentage of broilers slaughtered via CAS is rapidly increasing, from 20% in 2014 to 47% in 2021¹²⁵. It works by exposing broilers to either a mixture of inert gases (nitrogen and/or argon) or concentrations of carbon dioxide (CO₂), causing a reduction in available oxygen (0,) thus inducing loss of consciousness in the birds. In comparison to electrical water-bath stunning, one major advantage of CAS is that uncrating and shackling of live poultry can be completely eliminated, hence avoiding pre-slaughter handling-induced fear, anxiety, distress, suffering and pain in conscious birds^{130,134}. However, carbon dioxide is an acidic gas, causing the birds to experience some discomfort and stress before loss of consciousness if inhaled at high concentrations ^{129,135}. In multi-phase stunning systems, the birds are first exposed to relatively low concentrations of CO₂ (<40%) to be less aversive, and then, once the birds are unconscious, they are exposed to a higher concentration (80%-90%), which is sufficient to induce a deeper state of unconsciousness or death^{131,135}.

Low atmospheric pressure stunning (LAPS), a newly-approved method of killing poultry under EU law, kills birds with a slow, continuous, controlled decompression causing a gradual reduction of oxygen tension in the chamber, leading to progressive hypoxia^{136,137}. Loss of posture, regarded as a behavioural marker for loss of consciousness, occurs on average at 80 seconds¹³⁸. The major welfare benefits of LAPS over electrical water-bath stunning systems include no handling of live birds and no live shackling (since the birds are stunned in the modules used to transport them), no risk of pre-shocks, and no risk of ineffective stunning as LAPS reliably and irreversibly stuns all the birds^{137,139}. Some researchers consider LAPS to be superior than CAS since LAPS does not use any gases during the stunning process, being safer



for humans in the area¹⁴⁰ and because it is a less aversive method to the animals. There is also potential for small slaughterhouses to convert to this system more easily than to CAS. LAPS is not currently approved under the ECC due to limited research on the welfare implications of the slaughter method.

HIGHER WELFARE BREEDS

The use of higher welfare breeds accounts for less than 5% of the total number of broilers slaughtered in the EU⁹. However, higher welfare breeds represent 100% of the fresh retail market for chicken in the Netherlands¹⁴¹. In the UK, higher welfare breeds represent almost 5-11% of broiler production^{141,142,} while in France the Label Rouge higher welfare breeds make up around 15% of the market share¹⁴³.

The breeds currently acceptable for use under the RSPCA welfare standards for meat chicken include the JA757, 957, 787, 987, Norfolk Black, JACY57, Redbro, Ranger Classic, Rambler Ranger and the Ranger Gold. The most commonly-used breed is the JA787, while the JA757 remains their standard reference breed for which the RSPCA tests all other breeds against before entering them into the assurance scheme.

Slow-growing broilers have lower mortality rate than typical commercial fast-growing broilers^{65,144–146} and are less susceptible to leg disorders and heart problems^{65,146–149}. They are more active, perching, running and walking more than conventional breeds^{65,149–151}. Fast-growing broilers spend more time sitting on the floor, eating, and drinking than higher welfare birds^{65,152–155}. Although fast-growing broilers are motivated to perform their natural behaviours if their environment allows for this, their physical ability to perform some behaviours becomes increasingly restricted as they age, most probably because of their heavy weight^{151,156} and the high stocking densities observed in conventional production systems¹⁵⁷.

It is believed that selection for rapid growth reduces immune-competence and increases susceptibility to diseases¹⁵⁸. This has not only taken its toll on antibiotic usage across the industry, but the increased emergence of antibiotic resistant bacteria. The Food Standards Agency UK found that 60.6% of Campylobacter jejuni taken from retail chickens were resistant to tetracycline, an antibiotic widely used in human



medicine¹⁵⁹. Similarly, E. coli isolated from chickens in the US, Brazil, China, Poland, United Kingdom, Germany, France, and Spain were found to contain resistance to a number of antibiotics, including penicillins and tetracyclines¹⁶⁰. The breed impact is due to faster growing breeds requiring 3 to 9 times more antibiotics than slower growing breeds^{161–163}. This includes the use of fluoroquinolones, antibiotics considered "critically important in human medicine" whose "use in animals should be restricted to mitigate the risk to public health" ¹⁶⁴, in which usage was almost 6.7 times lower in slower growing breeds in 2021¹⁶⁵. This widespread adoption of higher welfare breeds in the Netherlands has allowed the overall industry antibiotic usage to drop by a third between 2017 and 2022. The majority of the farms raising slow-growing breeds (80%) did not record any antibiotic use for 2022¹⁶³. Alternatively, in the UK where this wide scale adoption of slower growing breeds has stalled, antibiotic usage actually increased by a third across the same period¹⁶⁶.

FIGURE 2: AVERAGE ANTIBIOTIC USE ON BROILERS IN THE NETHERLANDS FARMS -CONVENTIONAL VS ALTERNATIVE BREEDS, 2017-2022¹⁶³





WHY WE MUST ADDRESS ALL OF THESE ISSUES

While all of the factors addressed above have an impact on broiler welfare, all of them must be addressed for the maximum reduction in suffering. For example, switching to a slower growing breed but not addressing the other factors will result in birds that are more able to act out natural behaviours but do not have the space or enrichments to do so. Similarly, giving a faster growing breed more space does not address the breed susceptibility to death and disease or the other welfare concerns. Faster growing broilers at 30 kg/m² and with environmental enrichment have consistently higher gait scores, worse feather cover, are less reactive, have lower perch use, higher mortality and cull rates, and more downgraded meat, compared to slower growing birds under the same conditions¹⁴⁹. Combining breed and stocking density has the greatest benefits for broiler welfare¹⁶⁷.

Further demonstrating the importance of addressing all of the above issues, Schuck-Paim and Alonso¹⁶⁸ measured the impact of different production methods on the time spent in pain for the average animal within that system, directly comparing conventional production to production standards meeting the Better Chicken Commitment (BCC) requirements. They estimated that the adoption of the BCC resulted in a significant reduction in the amount of pain each animal endured, when compared to conventional production (table 2). <u>See here for full pain definitions.</u>

TABLE 2: THE REDUCTION IN PAIN WHEN SWITCHING FROM CONVENTIONAL PRODUCTION TO THE BCC¹⁶⁸

	Pain Reduction With the BCC	
	Before Slaughter	During Slaughter
Hurtful Disrupts the ability of individuals to function optimally	24%	
Disabling Pain that takes priority over most behaviours and prevents all forms of enjoyment or positive welfare	66%	87 - 90%
Excruciating Extreme levels of pain that are not normally tolerated even if only for a few seconds	78%	99 - 100%



COMMERCIAL VIABILITY

All of these welfare improvements can be made while remaining commercially viable and real world examples confirm this. Norsk Kylling is a poultry supplier in Norway that holds approximately 27% of the country's market share. They manage their whole supply chain, from parent flocks through to, and including, the processing plant. They committed to the European Chicken Commitment (ECC), which addresses all of the above welfare concerns, in 2020 and had fulfilled all of the requirements by spring 2022. Since making this change they now have; a 39% lower daily mortality, 79% lower DOA's (birds that die during transport), and an 80% lower incidence of ascites. Norsk Kylling now produces 3 million fewer birds each year, but still produces the same amount of meat¹⁶⁹.

CONCLUSION

The cost of cheap chicken is paid for with the suffering of fast-growing birds on a mass scale living in large and cramped sheds lacking in environmental stimulation, and experiencing poor slaughter practices. The impact of faster growth and enlarged breast muscle-driven by economic factors-is highlighted by the extensive list of diseases these birds are becoming increasingly predisposed to suffering.

However, there is an alternative; breeds exist that can alleviate many of the negative predispositions we see with the current typical fast-growing breeds. By utilising these higher welfare breeds and giving birds more space, enriching the environment, and improving slaughter conditions using multi-phase CAS, we would see an improvement in the level of welfare for the billions of chickens farmed for meat production every year.



REFERENCES

- European Commission-Directorate-General for Agriculture. Poultry Production. European Commission <u>https://agridata.ec.europa.eu/extensions/DashboardPoultry/PoultryProduction.</u> <u>html#</u> (2024).
- European Commission. EU agricultural outlook for markets, income and environment 2022-2032. https://doi.org/10.2762/29222 (2022).
- Agriculture and Horticulture Development Board. EU pork production at lowest level in almost a decade. <u>https://ahdb.org.uk/news/eu-pork-production-at-lowest-level-in-almost-a-decade</u> (2023).
- 4. EFSA Panel on Animal Health and Welfare. Welfare of broilers on farm. EFSA Journal 21, e07788 (2023).
- van Horne, P. L. M. Competitiveness of the EU Poultry Meat Sector, Base Year 2017; International Comparison of Production Costs. <u>http://dx.doi.org/10.18174/465696 (2018) doi:10.18174/465696.</u>
- 6. Zuidhof, M. J., Schneider, B. L., Carney, V. L., Korver, D. R. & Robinson, F. E. Growth, efficiency, and yield of commercial broilers from 1957, 1978, and 2005. Poultry Science 93, 2970–2982 (2014).
- European Commission. Report from the Commission to the European Parliament and the Council on the Impact of Genetic Selection on the Welfare of Chickens Kept for Meat Production. <u>eur-lex.europa.</u> <u>eu/legal-content/EN/TXT/PDF/?uri=CELEX:52016DC0182&from=EN</u> (2016).
- 8. Aviagen. Ross 308 Broiler Performance Objectives. <u>aviagen.com/assets/Tech_Center/Ross_Broiler/</u> <u>RossxRoss308-BroilerPerformanceObjectives2022-EN.pdf (</u>2022).
- European Commission. Study on the Application of the Broilers Directive (DIR 2007/43/EC) and Development of Welfare Indicators. <u>publications.europa.eu/en/publication-detail/-/publication/</u> <u>f4ccd35e-d004-11e7-a7df-01aa75ed71a1 (2017) doi:10.2875/729456 (2017).</u>
- Marino, L. Thinking chickens: a review of cognition, emotion, and behavior in the domestic chicken. Animal Cognition 20, 127–147 (2017).
- 11. Meluzzi, A. & Sirri, F. Welfare of broiler chickens. Italian Journal of Animal Science 8, 161–173 (2009).
- 12. Weeks, C. & Butterworth, A. Measuring and Auditing Broiler Welfare. (CABI Publishing, 2004).
- Pompeu, M. A. et al. Nutritional aspects related to non-infectious diseases in locomotor system of broilers. World's Poultry Science Journal 68, 669–678 (2012).
- 14. Wideman, R. F., Jr. Bacterial chondronecrosis with osteomyelitis and lameness in broilers: a review. Poultry Science 95, 325–344 (2016).
- 15. McGeown, D., Danbury, T. C., Waterman-Pearson, A. E. & Kestin, S. C. Effect of carprofen on lameness in broiler chickens. Veterinary Record 144, 668–671 (1999).
- Danbury, T. C., Weeks, C. A., Chambers, J. P., Waterman-Pearson, A. E. & Kestin, S. C. Self-selection of the analgesic drug carprofen by lame broiler chickens. Veterinary Record 146, 307–311 (2000).
- Knowles, T. G. et al. Leg disorders in broiler chickens: Prevalence, risk factors and prevention. PLoS One 3, 1–5 (2008).
- 18. Wijesurendra, D. S. et al. Pathological and microbiological investigations into cases of bacterial chondronecrosis and osteomyelitis in broiler poultry. Avian Pathology 46, 683–694 (2017).



- McNamee, P. T. & Smyth, J. A. Bacterial chondronecrosis with osteomyelitis ('femoral head necrosis') of broiler chickens: A review. Avian Pathology 29, 477–495 (2000).
- 20. Riddell, C. The development of tibial dyschondroplasia in broiler chickens. Avian Diseases 19, 443–462 (1975).
- 21. Leach, R. M., Jr & Monsonego-Ornan, E. Tibial dyschondroplasia 40 years later. Poultry Science 86, 2053–2058 (2007).
- 22. Van Der Heide, L. Viral arthritis/tenosynovitis: a review. Avian Pathology 6, 271-284 (1977).
- 23. Nham, E. G. et al. Flock-level prevalence, geographical distribution, and seasonal variation of avian reovirus among broiler flocks in Ontario. Canadian Veterinary Journal 58, 828–834 (2017).
- 24. Buijs, S., Keeling, L., Rettenbacher, S., van Poucke, E. & Tuyttens, F. A. M. Stocking density effects on broiler welfare: Identifying sensitive ranges for different indicators. Poultry Science 88, 1536–1543 (2009).
- De Jong, I. C., Gunnink, H. & van Harn, J. Wet litter not only induces footpad dermatitis but also reduces overall welfare, technical performance, and carcass yield in broiler chickens. Journal of Applied Poultry Research 23, 51–58 (2014).
- Julian, R. J. Rapid Growth Problems: Ascites and skeletal deformities in broilers. Poultry Science 77, 1773–1780 (1998).
- 27. Julian, R. J. Ascites in poultry. Avian Pathology 22, 419-454 (1993).
- 28. Bessei, W. Welfare of broilers: a review. World's Poultry Science Journal 62, 455 (2006).
- 29. Baghbanzadeh, A. & Decuypere, E. Ascites syndrome in broilers: Physiological and nutritional perspectives. Avian Pathology 37, 117–126 (2008).
- Wideman, R. F., Rhoads, D. D., Erf, G. F. & Anthony, N. B. Pulmonary arterial hypertension (ascites syndrome) in broilers: a review. Poultry Science 92, 64–83 (2013).
- Druyan, S. Ascites syndrome in broiler chickens A physiological syndrome affected by red blood cell. in Blood Cell - An Overview of Studies in Hematology (ed. Moschandreou, T.) 243–270 (InTech, 2012).
- 32. Balog, J. M. Ascites syndrome (Pulmonary Hypertension Syndrome) in broiler chickens: Are we seeing the light at the end of the tunnel? Avian and Poultry Biology Reviews 14, 99–126 (2003).
- Pavlidis, H. O. et al. Divergent selection for ascites incidence in chickens. Poultry Science 86, 2517–2529 (2007).
- 34. Part, C. E., Edwards, P., Hajat, S. & Collins, L. M. Prevalence rates of health and welfare conditions in broiler chickens change with weather in a temperate climate. Royal Society Open Science 3, 160197 (2016).
- 35. Forseth, M., Moe, R. O., Kittelsen, K., Skjerve, E. & Toftaker, I. Comparison of carcass condemnation causes in two broiler hybrids differing in growth rates. Scientific Reports 13, 4195 (2023).
- Ononiwu, J. C., Thomson, R. G., Carlson, H. C. & Julian, R. J. Studies on effect of lighting on 'Sudden death syndrome' in broiler chickens. Canadian Veterinary Journal 20, 74–77 (1979).
- Meshram, P. V. & Bijoy, F. Managemental and nutritional disease-sudden death syndrome in broilers. International Journal of Science, Environment and Technology 6, 260–266 (2017).
- Steele, P. & Edgar, J. Importance of acute death syndrome in mortalities in broiler chicken flocks. Australian Veterinary Journal 58, 63–66 (1982).
- Maxwell, M. H. & Robertson, G. W. UK survey of broiler ascites and sudden death syndromes in 1993. British Poultry Science 39, 203–215 (1998).



- Olkowski, A. A. & Classen, H. L. High incidence of cardiac arrhythmias in broiler chickens. Journal of Veterinary Medicine Series A 45, 83–91 (1998).
- Chung, H. C., Guenter, W., Rotter, R. G., Crow, G. H. & Stanger, N. E. Effects of dietary fat source on sudden death syndrome and cardiac sarcoplasmic reticular calcium transport in broiler chickens. Poultry Science 72, 310–316 (1993).
- 42. Olkowski, A. A. et al. A study on pathogenesis of sudden death syndrome in broiler chickens. Research in Veterinary Science 85, 131–140 (2008).
- 43. Olkowski, A. A. & Classen, H. L. Malignant ventricular dysrhythmia in broiler chickens dying of sudden death syndrome. Veterinary Record 140, 177–179 (1997).
- Gardiner, E. E., Hunt, J. R., Newberry, R. C. & Hall, J. W. Relationships between age, body weight, and season of the year and the incidence of sudden death syndrome in male broiler chickens. Poultry Science 67, 1243–1249 (1988).
- 45. Barbut, S. et al. Progress in reducing the pale, soft and exudative (PSE) problem in pork and poultry meat. Meat Science 79, 46–63 (2008).
- 46. Dransfield, E. & Sosnicki, A. A. Relationship between muscle growth and poultry meat quality. Poultry Science 78, 743–746 (1999).
- 47. Mudalal, S., Lorenzi, M., Soglia, F., Cavani, C. & Petracci, M. Implications of white striping and wooden breast abnormalities on quality traits of raw and marinated chicken meat. Animal 9, 728–734 (2015).
- 48. Maiorano, G. Meat defects and emergent muscle myopathies in broiler chickens: Implications for the modern poultry industry. Scientific Annals of Polish Society of Animal Production 13, 43–51 (2017).
- 49. Norring, M. et al. Wooden breast myopathy links with poorer gait in broiler chickens. Animal 1-6 (2018).
- 50. Huang, X. & Ahn, D. U. The incidence of muscle abnormalities in broiler breast meat A review. Korean Journal for Food Science of Animal Resources 38, 835–850 (2018).
- 51. Kuttappan, V. A., Hargis, B. M. & Owens, C. M. White striping and woody breast myopathies in the modern poultry industry: a review. Poultry Science 95, 2724–2733 (2016).
- 52. Aviagen Meat Quality Working Group. Breast Muscle Myopathies (BMM). <u>en.aviagen.com/assets/</u> Tech_Center/Broiler_Breeder_Tech_Articles/English/Breast-Muscle-Myopathies-2019-EN.pdf (2019).
- 53. Bilgili, J. & Hess, J. Green muscle disease. Reducing the incidence in broiler flock. Ross Tech vol. 8 hen.aviagen.com/assets/Tech_Center/AA_Technical_Articles/AAUpdateGreenMuscle.pdf (2008).
- 54. Kijowski, J., Kupińska, E., Stangierski, J., Tomaszewska-Gras, J. & Szablewski, T. Paradigm of deep pectoral myopathy in broiler chickens. World's Poultry Science Journal 70, 125–138 (2014).
- Dinev, I. & Kanakov, D. Deep pectoral myopathy: Prevalence in 7 weeks old broiler chickens in Bulgaria. Revue de médecine vétérinaire 162, 279–283 (2011).
- Lien, R. J., Bilgili, S. F., Hess, J. B. & Joiner, K. S. Induction of deep pectoral myopathy in broiler chickens via encouraged wing flapping. Journal of Applied Poultry Research 21, 556–562 (2012).
- Papah, M. B., Brannick, E. M., Schmidt, C. J. & Abasht, B. Evidence and role of phlebitis and lipid infiltration in the onset and pathogenesis of Wooden Breast Disease in modern broiler chickens. Avian Pathology 46, 623–643 (2017).
- Sihvo, H.-K. et al. Wooden breast myodegeneration of pectoralis major muscle over the growth period in broilers. Veterinary Pathology 54, 119–128 (2017).



- Petracci, M., Mudalal, S., Bonfiglio, A. & Cavani, C. Occurrence of white striping under commercial conditions and its impact on breast meat quality in broiler chickens. Poultry Science 92, 1670– 1675 (2013).
- 60. Bailey, R. A., Watson, K. A., Bilgili, S. F. & Avendano, S. The genetic basis of pectoralis major myopathies in modern broiler chicken lines. Poultry Science 94, 2870–2879 (2015).
- 61. Zambonelli, P. et al. Detection of differentially expressed genes in broiler pectoralis major muscle affected by White Striping Wooden Breast myopathies. Poultry Science 95, 2771–2785 (2016).
- 62. Abasht, B., Mutryn, M. F., Michalek, R. D. & Lee, W. R. Oxidative stress and metabolic perturbations in wooden breast disorder in chickens. PLoS One 11, 1–16 (2016).
- Mutryn, M. F., Brannick, E. M., Fu, W., Lee, W. R. & Abasht, B. Characterization of a novel chicken muscle disorder through differential gene expression and pathway analysis using RNA-sequencing. BMC Genomics 16, 399 (2015).
- 64. Kuttappan, V. A. et al. Estimation of factors associated with the occurrence of white striping in broiler breast fillets. Poultry Science 92, 811–819 (2013).
- 65. Dixon, L. M. Slow and steady wins the race: The behaviour and welfare of commercial faster growing broiler breeds compared to a commercial slower growing breed. PLoS One 15, e0231006 (2020).
- Zebracki, K. & Drotar, D. Pain and activity limitations in children with Duchenne or Becker muscular dystrophy. Developmental Medicine & Child Neurology 50, 546–552 (2008).
- 67. Silva, T. D. da et al. Pain characterization in Duchenne muscular dystrophy. Arquivos de Neuropsiquiatria 74, 767–774 (2016).
- 68. Soglia, F. et al. Histology, composition, and quality traits of chicken Pectoralis major muscle affected by wooden breast abnormality. Poultry Science 95, 651–659 (2016).
- 69. Kuttappan, V. A., Brewer, V. B., Apple, J. K., Waldroup, P. W. & Owens, C. M. Influence of growth rate on the occurrence of white striping in broiler breast fillets. Poultry Science 91, 2677–2685 (2012).
- 70. Baldi, G. et al. Implications of white striping and spaghetti meat abnormalities on meat quality and histological features in broilers. Animal 12, 164–173 (2018).
- 71. European Commission. Report from the Commission to the European Parliament and the Council on the Application of Directive 2007/43/EC and Its Influence on the Welfare of Chickens Kept for Meat Production, as Well as the Development of Welfare Indicators. <u>eur-lex.europa.eu/legal-content/EN/ TXT/PDF/?uri=CELEX:52018DC0181&from=EN</u> (2018).
- Spindler, B., Briese, A. & Hartung, J. How much floor space needs a broiler chicken? in Proceedings of the XV International Congress of the International Society for Animal Hygiene (2011). doi:10.13140/2.1.4485.0240.
- Zhao, F., Zhao, Y., Geng, A., Shi, Z. & Li, B. Effects of stocking density on behavior of broilers in cage system. in Proceedings of the VIII Livestock Environment Symposium (2008). doi:10.13031/2013.25553.
- 74. Dawkins, M. S., Donnelly, C. A. & Jones, T. A. Chicken welfare is influenced more by housing conditions than by stocking density. Nature 427, 342–344 (2004).
- 75. Dawkins, M. S. Stocking density: Can we judge how much space poultry need? In Advances in Poultry Welfare. (Ed. Mench. J.) 227–242. Woodhead Publishing (2018).
- Yakubu, A., Gwaska, J. A. & Salako, A. E. Strain and placement density effects on welfare, haematological and serum biochemical indices of broilers in north central Nigeria. Acta Agriculturae Slovenica 94, 153–158 (2009).



- 77. Feddes, J. J., Emmanuel, E. J. & Zuidhoft, M. J. Broiler performance, body weight variance, feed and water intake, and carcass quality at different stocking densities. Poultry Science 81, 774–779 (2002).
- 78. Kryeziu, A. J., Kamberi, M., Muji, S. & Mestani, N. Carcass traits of broilers as affected by different stocking density and sex. Bulgarian Journal of Agricultural Science 24, 1097–1103 (2018).
- 79. Dozier, W. A., III et al. Stocking density effects on growth performance and processing yields of heavy broilers. Poultry Science 84, 1332–1338 (2005).
- 80. Elfadil, A. A., Vaillancourt, J. P. & Meek, A. H. Impact of stocking density, breed, and feathering on the prevalence of abdominal skin scratches in broiler chickens. Avian Diseases 40, 546–552 (1996).
- Skrbic, Z. et al. The effect of stocking density on individual broiler welfare parameters: 2. Different broiler stocking densities. Biotechnology in Animal Husbandry 27, 17–24 (2011).
- Bailie, C. L., Ijichi, C. & O'Connell, N. E. Effects of stocking density and string provision on welfarerelated measures in commercial broiler chickens in windowed houses. Poultry Science 97, 1503– 1510 (2018).
- 83. Singh, K. D., Pramanik, P. S. & Kashyap, S. S. Effect of stocking density on foot pad lesions and mortality in broiler chickens. Bulletin of Environment, Pharmacology and Life Sciences 6, 19–24 (2017).
- Sørensen, P., Su, G. & Kestin, S. C. Effects of age and stocking density on leg weakness in broiler chickens. Poultry Science 79, 864–870 (2000).
- 85. Kristensen, H. H. & Wathes, C. M. Ammonia and poultry welfare: a review. World's Poultry Science Journal 56, 235–245 (2000).
- Wei, F. X. et al. Ammonia concentration and relative humidity in poultry houses affect the immune response of broilers. Genetics and Molecular Research 14, 3160–3169 (2015).
- 87. Olanrewaju, H. A., Thaxton, J. P., Dozier, W. A., III, Purswell, J. & Branton, S. L. A review of lighting programs for broiler production. International Journal of Poultry Science 5, (2006).
- Schwean-Lardner, K., Fancher, B. I., Laarveld, B. & Classen, H. L. Effect of day length on flock behavioural patterns and melatonin rhythms in broilers. British Poultry Science 55, 21–30 (2014).
- 89. COUNCIL DIRECTIVE 2007/43/EC. Official Journal of the European Union <u>eur-lex.europa.eu/</u> LexUriServ/LexUriServ.do?uri=OJ:L:2007:182:0019:0028:EN:PDF (2007).
- Kristensen, H. H., Aerts, J. M., Leroy, T., Wathes, C. M. & Berckmans, D. Modelling the dynamic activity of broiler chickens in response to step-wise changes in light intensity. Applied Animal Behaviour Science 101, 125–143 (2006).
- Rault, J.-L., Clark, K., Groves, P. J. & Cronin, G. M. Light intensity of 5 or 20 lux on broiler behavior, welfare and productivity. Poultry Science 96, 779–787 (2017).
- Alvino, G. M., Archer, G. S. & Mench, J. A. Behavioural time budgets of broiler chickens reared in varying light intensities. Applied Animal Behaviour Science 118, 54–61 (2009).
- Deep, A., Schwean-Lardner, K., Crowe, T. G., Fancher, B. I. & Classen, H. L. Effect of light intensity on broiler behaviour and diurnal rhythms. Applied Animal Behaviour Science 136, 50–56 (2012).
- 94. Mendes, A. S. et al. Performance and preference of broiler chickens exposed to different lighting sources. Journal of Applied Poultry Research 22, 62–70 (2013).
- 95. Newberry, R. C., Hunt, J. R. & Gardiner, E. E. Influence of light intensity on behavior and performance of broiler chickens. Poultry Science 67, 1020–1025 (1988).



- Harrison, P. C., Bercovitz, A. B. & Leary, G. A. Development of eye enlargement of domestic fowl subjected to low intensity light. International Journal of Biometeorology 12, 351–358 (1968).
- Deep, A., Raginski, C., Schwean-Lardner, K., Fancher, B. I. & Classen, H. L. Minimum light intensity threshold to prevent negative effects on broiler production and welfare. British Poultry Science 54, 686–694 (2013).
- Davis, N. J., Prescott, N. B., Savory, C. J. & Wathes, C. M. Preferences of growing fowls for different light intensities in relation to age, strain and behaviour. Animal Welfare 8, 193–203 (1999).
- 99. Blatchford, R. A. et al. The effect of light intensity on the behavior, eye and leg health, and immune function of broiler chickens. Poultry Science 88, 20–28 (2009).
- 100. Bayram, A. & Özkan, S. Effects of a 16-hour light, 8-hour dark lighting schedule on behavioral traits and performance in male broiler chickens. Journal of Applied Poultry Research 19, 263–273 (2010).
- 101. Schwean-Lardner, K., Fancher, B. I. & Classen, H. L. Impact of daylength on behavioural output in commercial broilers. Applied Animal Behaviour Science 137, 43–52 (2012).
- 102. Sanotra, G. S., Lund, J. D. & Vestergaard, K. S. Influence of light-dark schedules and stocking density on behaviour, risk of leg problems and occurrence of chronic fear in broilers. British Poultry Science 43, 344–354 (2002).
- 103. Zulkifli, I., Rasedee, A., Syaadoh, O. N. & Norma, M. T. C. Daylength effects on stress and fear responses in broiler chickens. Asian-Australasian Journal of Animal Sciences 11, 751–754 (1998).
- 104. E. E. Onbaşılar, E. E., Erol, H., Cantekin, Z. & Kaya, Ü. Influence of intermittent lighting on broiler performance, incidence of tibial dyschondroplasia, tonic immobility, some blood parameters and antibody production. Asian-Australasian Association of Animal Production Societies 20, 550–555 (2007).
- 105. Li, T., Howland, H. C. & Troilo, D. Diurnal illumination patterns affect the development of the chick eye. Vision Research 40, 2387–2393 (2000).
- 106. Schwean-Lardner, K., Fancher, B. I. & Classen, H. L. Impact of daylength on the productivity of two commercial broiler strains. British Poultry Science 53, 7–18 (2012).
- 107. Schwean-Lardner, K. et al. Effect of day length on cause of mortality, leg health, and ocular health in broilers. Poultry Science 92, 1–11 (2013).
- 108. Hassanzadeh, M., Bozorgmerifard, M. H., Akbari, A. R., Buyse, J. & Decuypere, E. Effect of intermittent lighting schedules during the natural scotoperiod on T3-induced ascites in broiler chickens. Avian Pathology 29, 433–439 (2000).
- 109. Zheng, L. et al. Growth performance, antioxidant status, and nonspecific immunity in broilers under different lighting regimens. Journal of Applied Poultry Research 22, 798–807 (2013).
- 110. Bailie, C. L., Ball, M. E. E. & O'Connell, N. E. Influence of the provision of natural light and straw bales on activity levels and leg health in commercial broiler chickens. Animal 7, 618–626 (2013).
- 111. Sans, E. C. de O. et al. From the point of view of the chickens: what difference does a window make? Animals (Basel) 11, (2021).
- 112. Illuminance Recommended Light Levels. Engineeringtoolbox. <u>engineeringtoolbox.com/light-level-</u> rooms-d_708.html.
- 113. Newberry, R. C. Environmental enrichment: Increasing the biological relevance of captive environments. Applied Animal Behaviour Science 44, 229–243 (1995).



- 114. Riber, A. B., van de Weerd, H. A., de Jong, I. C. & Steenfeldt, S. Review of environmental enrichment for broiler chickens. Poultry Science 97, 378–396 (2018).
- 115. Lourenço da Silva, M. I. et al. Behaviour and animal welfare indicators of broiler chickens housed in an enriched environment. PLoS One 16, e0256963 (2021).
- 116. Vasdal, G., Vas, J., Newberry, R. C. & Moe, R. O. Effects of environmental enrichment on activity and lameness in commercial broiler production. Journal of Applied Animal Welfare Science 22, 197–205 (2019).
- 117. Rodriguez-Aurrekoetxea, A., Leone, E. H. & Estevez, I. Effects of panels and perches on the behaviour of commercial slow-growing free-range meat chickens. Applied Animal Behaviour Science 165, 103–111 (2015).
- 118. Ventura, B. A., Siewerdt, F. & Estevez, I. Access to barrier perches improves behavior repertoire in broilers. PLoS One 7, e29826 (2012).
- 119. Zhao, J. P. et al. Cool perches improve the growth performance and welfare status of broiler chickens reared at different stocking densities and high temperatures. Poultry Science 92, 1962–1971 (2013).
- 120. Ventura, B. A., Siewerdt, F. & Estevez, I. Effects of barrier perches and density on broiler leg health, fear, and performance. Poultry Science 89, 1574–1583 (2010).
- 121. Kaukonen, E., Norring, M. & Valros, A. Perches and elevated platforms in commercial broiler farms: use and effect on walking ability, incidence of tibial dyschondroplasia and bone mineral content. Animal 11, 864–871 (2017).
- 122. Cornetto, T., Estevez, I. & Douglass, L. W. Using artificial cover to reduce aggression and disturbances in domestic fowl. Applied Animal Behaviour Science 75, 325–336 (2002).
- 123. Cornetto, T. & Estevez, I. Behavior of the domestic fowl in the presence of vertical panels. Poultry Science 80, 1455–1462 (2001).
- 124. Bailie, C. L. & O'Connell, N. E. The influence of providing perches and string on activity levels, fearfulness and leg health in commercial broiler chickens. Animal 9, 660–668 (2015).
- 125. European Union Reference Centre for Animal Welfare. Main welfare aspects of stunning broilers by exposure to controlled atmosphere. <u>sitesv2.anses.fr/en/system/files/EURCAW-Poultry-SFA_Factsheet_CAS.pdf</u> (2022).
- 126. Hindle, V. A., Lambooij, E., Reimert, H. G. M., Workel, L. D. & Gerritzen, M. A. Animal welfare concerns during the use of the water bath for stunning broilers, hens, and ducks. Poultry Science 89, 401–412 (2010).
- 127. Gentle, M. J. & Tilston, V. L. Nociceptors in the legs of poultry: Implications for potential pain in pre-slaughter shackling. Animal Welfare 9, 227–236 (2000).
- 128. Bedanova, I. et al. Stress in broilers resulting from shackling. Poultry Science 86, 1065–1069 (2007).
- 129. Raj, M. Welfare during stunning and slaughter of poultry. Poultry Science 77, 1815–1819 (1998).
- 130. Shields, S. J. & Raj, A. B. M. A critical review of electrical water-bath stun systems for poultry slaughter and recent developments in alternative technologies. Journal of Applied Animal Welfare Science 13, 281–299 (2010).
- 131. Berg, C. & Raj, M. A review of different stunning methods for poultry–animal welfare aspects (stunning methods for poultry). Animals 5, 1207–1219 (2015).



- 132. Raj, M. & Tsemeni-Gousi, A. Stunning methods for poultry. World's Poultry Science Journal 56, 291–304 (1996).
- 133. Devos, G., Moons, C. P. H. & Houf, K. Diversity, not uniformity: slaughter and electrical waterbath stunning procedures in Belgian slaughterhouses. Poultry Science 97, 3369–3379 (2018).
- 134. Raj, A. B. M. Recent developments in stunning and slaughter of poultry. World's Poultry Science Journal 62, 467–484 (2006).
- 135. Gerritzen, M. A., Reimert, H. G. M., Hindle, V. A., Verhoeven, M. T. W. & Veerkamp, W. B. Multistage carbon dioxide gas stunning of broilers. Poultry Science 92, 41–50 (2013).
- 136. EFSA Panel on Health and Welfare. Low atmospheric pressure system for stunning broiler chickens. EFSA Journal 15, e05056 (2017).
- 137. McKeegan, D. E. F., Sandercock, D. A. & Gerritzen, M. A. Physiological responses to low atmospheric pressure stunning and the implications for welfare. Poultry Science 92, 858–868 (2013).
- 138. Mackie, N. & McKeegan, D. E. F. Behavioural responses of broiler chickens during low atmospheric pressure stunning. Applied Animal Behaviour Science 174, 90–98 (2016).
- Martin, J. E., Christensen, K., Vizzier-Thaxton, Y. & McKeegan, D. E. F. Effects of analgesic intervention on behavioural responses to Low Atmospheric Pressure Stunning. Applied Animal Behaviour Science 180, 157–165 (2016).
- 140. Vizzier-Thaxton, Y., Christensen, K. D., Schilling, M. W., Buhr, R. J. & Thaxton, J. P. A new humane method of stunning broilers using low atmospheric pressure. Journal of Applied Poultry Research 19, 341–348 (2010).
- 141. Compassion in World Farming. Information Sheet 1 Chicken meat production in the EU & UK. compassioninfoodbusiness.com/media/7455892/info-sheet-1-broiler-production-eu-and-uk.pdf.
- 142. Mcdougal, T. Shifting UK poultry meat towards slower-growing breeds. Poultry World (2023).
- 143. Compassion in World Farming. Chicken Track: Rapport Europeen 2022. <u>ciwf.org.uk/media/7453679/</u> <u>ciwf-rapport-europeen-chickentrack-2022.pdf</u> (2022).
- 144. Castellini, C., Dal Bosco, A., Mugnai, C. & Bernardini, M. Performance and behaviour of chickens with different growing rate reared according to the organic system. Italian Journal of Animal Science 1, 291–300 (2002).
- 145. Fanatico, A. C. et al. Performance, livability, and carcass yield of slow- and fast-growing chicken genotypes fed low-nutrient or standard diets and raised indoors or with outdoor access. Poultry Science 87, 1012–1021 (2008).
- 146. Rayner, A. C., Newberry, R. C., Vas, J. & Mullan, S. Slow-growing broilers are healthier and express more behavioural indicators of positive welfare. Scientific Reports 10, 15151 (2020).
- 147. Wilhelmsson, S. Comparison of behaviour and health of two broiler hybrids with different growth rates. (SLU, Dept. of Animal Environment and Health, 2016).
- 148. Abeyesinghe, S. M. et al. Associations between behaviour and health outcomes in conventional and slow-growing breeds of broiler chicken. Animal 15, 100261 (2021).
- 149. Baxter, M., Richmond, A., Lavery, U. & O'Connell, N. E. A comparison of fast growing broiler chickens with a slower-growing breed type reared on Higher Welfare commercial farms. PLoS One 16, e0259333 (2021).



- 150. Dawson, L. C., Widowski, T. M., Liu, Z., Edwards, A. M. & Torrey, S. In pursuit of a better broiler: a comparison of the inactivity, behavior, and enrichment use of fast- and slower growing broiler chickens. Poultry Science 100, 101451 (2021).
- 151. Bokkers, E. A. M. & Koene, P. Behaviour of fast- and slow growing broilers to 12 weeks of age and the physical consequences. Applied Animal Behaviour Science 81, 59–72 (2003).
- 152. Nielsen, B. L., Juul-Madsen, H. R., Steenfeldt, S., Kjaer, J. B. & Sørensen, P. Feeding activity in groups of newly hatched broiler chicks: effects of strain and hatching time. Poultry Science 89, 1336–1344 (2010).
- 153. Dal Bosco, A., Mugnai, C., Sirri, F., Zamparini, C. & Castellini, C. Assessment of a global positioning system to evaluate activities of organic chickens at pasture. Journal of Applied Poultry Research 19, 213–218 (2010).
- 154. Fanatico, A. C., Owens, C. M. & Emmert, J. L. Organic poultry production in the United States: Broilers. Journal of Applied Poultry Research 18, 355–366 (2009).
- 155. de Jong, I. C. et al. Providing environmental enrichments affects activity and performance, but not leg health in fast- and slower-growing broiler chickens. Applied Animal Behaviour Science 241, 105375 (2021).
- 156. Bokkers, E. A. M., Zimmerman, P. H., Bas Rodenburg, T. & Koene, P. Walking behaviour of heavy and light broilers in an operant runway test with varying durations of feed deprivation and feed access. Applied Animal Behaviour Science 108, 129–142 (2007).
- 157. Bergmann, S. et al. Behavior as welfare indicator for the rearing of broilers in an enriched husbandry environment–A field study. Journal of Veterinary Behavior 19, 90–101 (2017).
- 158. Swaggerty, C. L. et al. Selection of broilers with improved innate immune responsiveness to reduce on-farm infection by foodborne pathogens. Foodborne Pathogens and Disease 6, 777–783 (2009).
- 159. Food Standards Agency. Enhanced Molecular-Based Surveillance and Source Attribution of Campylobacter Infections in the UK. <u>food.gov.uk/print/pdf/node/6946</u> (2021) doi:10.46756/sci. fsa.ksj135.
- 160. Roth, N. et al. The application of antibiotics in broiler production and the resulting antibiotic resistance in Escherichia coli: A global overview. Poultry Science 98, 1791–1804 (2019).
- 161. Speksnijder, D. C., Mevius, D. J., Bruschke, C. J. M. & Wagenaar, J. A. Reduction of veterinary antimicrobial use in the Netherlands. The Dutch success model. Zoonoses Public Health 62 Suppl 1, 79–87 (2015).
- 162. Alliance to Save our Antibiotics. Antibiotic Use in the UK Poultry Sector. <u>saveourantibiotics.org/</u> media/1763/antibiotic-use-in-the-uk-poultry-sector.pdf (2016).
- 163. Netherlands Veterinary Medicines Institute (SDa). Usage of Antibiotics in Agricultural Livestock in the Netherlands in 2022. <u>cdn.i-pulse.nl/autoriteitdiergeneesmiddelen/userfiles/EN/SDa-rapporten/</u> <u>engels-def-sda-rapport-met-brief---het-gebruik-van-antibiotica-bij-landbouwhuisdieren-in-2022-</u> <u>revision.pdf</u> (2023).
- 164. European Medicines Agency. Categorisation of antibiotics used in animals promotes responsible use to protect public and animal health. <u>ema.europa.eu/en/news/categorisation-antibiotics-used-animals-promotes-responsible-use-protect-public-and-animal-health</u> (2020).
- 165. AVINED. Antibioticumgebruik Pluimveesector in 2021 en de trends van de afgelopen jaren. <u>avined.nl/</u> wp-content/uploads/2022-091-E0002-Infographic-2021-V006.pdf (2022).



- 166. UK-VARSS. Supplementary Material 2 (UK-VARSS 2021). <u>assets.publishing.service.gov.uk/</u> media/63a420188fa8f539169b1d93/TP_FOR_PUBLICATION_-_UK-VARSS_2021_Supplementary_ <u>Material_2-accessible.pdf</u> (2022).
- 167. van der Eijk, J. A. J. et al. Fast- and slower-growing broilers respond similarly to a reduction in stocking density with regard to gait, hock burn, skin lesions, cleanliness, and performance. Poultry Science 102, 102603 (2023).
- 168. Schuck-Paim, C. & Alonso, W. J. Quantifying Pain in Broiler Chickens. (2022).
- 169. Norsk-Kylling. Responsibility Report. <u>files-cdn.vitaminw.no/18df4859adda4b9dbe14bffb7ab5f8ed/</u> <u>Root/Rapporter/221101+Responsibility+report+Norsk+Kylling+2021.pdf</u> (2021).